RecombiLISA

HBc IgM ELISA

IVD REF E0811

- 96-well ELISA Test for the qualitative detection of anti-hepatitis B virus core antigen (HBc) IgM in human serum or plasma
- . For export only, not for re-sale in the USA
- Store at 2-8°C upon receipt

INTENDED USE

The RecombiLISA HBc IgM ELISA is a solid-phase enzyme-linked immunosorbent assay for the qualitative detection of IgM anti-hepatitis B virus core antigen (HBc) in human serum or plasma. It is intended for professional use only as an aid in the diagnosis of infection with HBV.

INTRODUCTION

Hepatitis virus B (HBV) is the most common cause of persistent viremia and the most important cause of chronic liver disease and hepatocellular carcinoma. Clinically apparent HBV infections may have been extant for several millennia. It is estimated that there are 300 million chronic carriers of HBV in the world. The carrier rates vary from as little as 0.3% (Western countries) to 20% (Asia, Africa).

HBV is a hepatotropic DNA virus. The core of the virus contains a DNA polymerase², the core antigen (HBcAg)³ and the e antigen (HBeAg)⁴. The core of HBV is enclosed in a coat that contains lipid, protein and carbohydrate and expresses an antigen terms hepatitis B surface antigen (HBsAg)³.

Antibody to HBcAg (anti-HBc) appears shortly after HBsAg and before the appearance of detectable antibody to HBsAg, roughly at the time that serum ALT begins to rise. Anti-HBc also remains elevated for life and is a useful marker of the ongoing HBV infection as HBcAg itself does not circulate freely in the serum of such infected persons 1.4.

TEST PRINCIPLE

The RecombiLISA HBc IgM ELISA is a solid-phase enzyme-linked immunosorbent assay based on the principle of the IgM capture technique for the detection of IgM antibody to HBc in human serum or plasma.

The RecombiLISA HBc IgM ELISA is composed of two key components:

- 1) Solid microwells pre-coated with monoclonal anti-human IgM antibody
- 2) Liquid conjugates composed of HBc antigen conjugated with horseradish peroxidase (HRP-HBcAg Conjugate)

During the assay, the test specimen is first incubated in the coated microwells. Anti-HBc IgM, if present in the test specimen, binds to the antibody coated on the microwell surface. Unbound specimen is then removed by washing.

In the second incubation with the HRP-HBc Ag Conjugate, the anti-HBc IgM absorbed on the surface of microwell surface binds to the HBc Ag in the conjugate, forming a conjugate complex.

Unbound conjugate is then removed by washing. The presence of the conjugate complex is shown by the presence of a blue color upon additional incubation with TMB substrate. The reaction is stopped with Stop Solution and absorbances are read using a spectrophotometer at 450 /620-690 nm.

MATERIALS AND REAGENTS

Materials and reagents provided with the kit

ltem	Description	Quantity	Catalog			
1	Anti-human IgM Coated Microwells	12 wells x 8 strips	E0811W			
2	Sample Diluent (10x Concentrated)	10 mL	E0811SD			
3	HBc IgM Negative Control	1 mL	E0811N			
4	HBc IgM Positive Control	1 mL	E0811P			
5	HRP-HBcAg Conjugate	6 mL	E0811H			
6	Wash Buffer (30 x Concentrate)	20 mL	WE3000			
7	TMB Substrate A	6 mL	TME2000A			
8	TMB Substrate B	6 mL	TME2000B			
9	Stop Solution	6 mL	SE1000			
10	ELISA Working Sheet	2	E0001ES			
11	Product Insert	1	PI-E0811			
Others	ers 3 x Microplate Sealers and 1 x Resealable Bag					

Materials and reagents required but not provided in the kit

- Pipette capable of delivering 1 uL, 50 μL, 100 μL, and 1000 μL volumes
- Test tubes for specimen dilution

- Microplate reader with a bandwidth of 10 nm or less and an optical density range of 0-3OD or greater at 450nm wavelength is acceptable
- 4. Absorbent paper for blotting the microwells
- Time
- Distilled or de-ionized water

STORAGE AND STABILITY

All reagents except the concentrated wash buffer are ready to use as supplied. Store all components at 2-8°C. Do not freeze. Avoid strong light. Ensure that the reagents are brought to room temperature before opening. Reseal the microwells after removing the desired number of wells. Place unused wells in the resealable plastic bag provided and return to 2-8°C. All reagents are stable through the expiration date printed on the label if not opened.

SPECIMEN COLLECTION AND PREPARATION

- Serum or plasma should be prepared from a whole blood specimen obtained by acceptable venipuncture technique.
- This kit is designed for use with serum or plasma specimen without additives only.
- If a specimen is not tested immediately, refrigerated at 2°C-8°C. If storage
 period greater than three days are anticipated, the specimen should be frozen
 (-20°C). Avoid repeated freezing-thawing of specimens. If a specimen is to be
 shipped, pack in compliance with federal regulation covering the
 transportation of etiologic agents.
- Specimens containing precipitants may give inconsistent test results. Clarify such specimens by centrifugation prior to assaying.
- Do not use serum specimens demonstrating gross lipemia, gross hemolysis or turbidity. Do not use specimens containing sodium azide.

PREPARATION OF THE REAGENTS

Bring all reagents, controls to room temperature (18°C-28°C).

Preparation of working Wash Buffer:

If precipitants are visible, warm up the Wash Buffer (30X concentrate) at 37°C. Dilute concentrated Wash Buffer 30 fold with water as follows:

Plate	DI water	Wash buffer (30X)	Final volume
1 strip	58 mL	2.0 mL	60 mL
2 strips	116 mL	4.0 mL	120 mL
3 strips	174 mL	6.0 mL	180 mL
4 strips	232 mL	8.0 mL	240 mL

- Concentrated Sample Diluent contains 9% NaCl. Dilute it with water prior to use as following: Add 10 mL of the concentrated Sample Diluent into 90 mL of water and mix well. The working sample diluent contains 0.9% NaCl. Alternatively, user can use Saline Buffer as the sample diluent.
- 4. Dilute test specimen with the working sample diluent at 1:1000 dilution, ie: 1 µL of test serum or plasma into 1000 µL of the working sample diluent.
- Determine the number of microwells needed and mark on the ELISA Working Sheet with the appropriate information. Positive and Negative Controls require to be run in duplicate to ensure accuracy.
- Mix each reagent before adding to the test wells.

ASSAY PROCEDURE

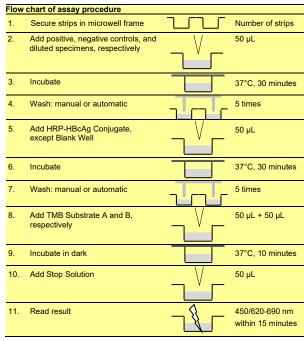
- Calculate the desired number of microwells. Remove the remaining microwells and place them with desiccant into the resealable plastic bag, seal and store at 2-8°C for later use.
- 2. Add specimens according to the designation on the ELISA Working Sheet:
- 2.1 Blank Well: Do not add any reagents.
- 2.2 <u>Control Wells:</u> Add 50 µL of HBc IgM Positive, Negative Control into the designated control wells, respectively.
- 2.3 Test Wells: Add 50 μL of diluted test specimens into each test well, respectively.
- Incubate the wells at 37°C for 30 minutes.
- 4. Wash Step (Can be performed manually or with automated washing):

Manual washing: Carefully remove the incubation mixture by disposing the solution into a waste container. Fill each well with 350 µL diluted wash buffer and rock gently for 20-30 seconds. Discard the wash solution completely. Repeat 4 more times. After completing the last wash step, tap the plate on absorbent paper to remove residual liquid.

Automatic washing: Automatic plate washer must be calibrated to ensure efficient washing. Aspirate incubation mixture from all wells completely. Fill each well with 350 uL diluted wash buffer and soak for 20-30 seconds.

Aspirate all wells completely. Repeat 4 more times

- 5. Add 50 µL of HRP-HBcAg Conjugate into each well except the Blank Well.
- 6. Cover the microwells and incubate at 37°C for 30 minutes.
- 7. Wash the plate 5 times as step 4 described.
- Add 50 of TMB Substrate A and 50 µL of TMB Substrate B into each well including the Blank Well.
- Incubate at 37°C in dark for 10 minutes.
- Stop the reaction by adding 50 µL of Stop Solution to each well. Gently mix for 30 seconds. It is important to make sure that all the blue color changes to yellow color completely.
- 11. Set the microplate reader wavelength at 450 nm and measure the absorbance (OD) of each well against the blank well within 15 minutes after adding Stop Solution. A filter of 620-690 nm can be used as a reference wavelength to optimize the assay result.



INTERPRETATION OF RESULTS

A. Set up the cut-off value

The cut-off value = $N \times 2.1$

N: Mean OD of the negative control. Use 0.05 for calculation of the cut-off value if the mean OD is less than 0.05.

B. Calculation of specimen OD ratio

Calculate an OD ratio for each specimen by dividing its OD value by the cutoff value as follows:

Specimen OD ratio = Specimen OD Cut-off Value

C. Assay validation

The mean OD value of the HBc IgM positive controls should be \geq 0.80. The mean OD value of the HBc IgM negative controls should be \leq 0.10.

Check the procedure and repeat assay if above conditions are not met.

D. Interpretation of the results

Specimen OD ratio

Negative < 1.00

- The negative result indicates that there is no detectable anti-HBc IgM in the specimen.
- Specimens with OD ratio ≥ 1.00 are initially considered to be positive by the RecombiLISA HBc IgM ELISA. They should be retested in duplicate before final interpretation.
- Results just below the cut-off value (Lower than 10% of the cut-off value) should be interpreted with caution (it is advisable to retest in duplicate the corresponding specimens when it is applicable).

If after retesting the absorbance of one of the duplicates is equal or greater than the cut-off value, the initial result is repeatable and the specimen is considered to be positive with the *RecombiLISA* HBc IgM ELISA, subject to the limitation of the procedure, described below.

If after re-testing of a specimen, the absorbance value of the 2 duplicates are less than the cut-off value, the initial result is non-repeatable and the specimen is considered to be negative with the RecombiLISA HBc IgM ELISA. Non-repeatable reactions are often caused by:

- Inadequate microwell washing
- Contamination of negative specimens by serum or plasma with a high antibody titer
- Contamination of the substrate solution by oxidizing agents (bleach, metal ions, etc.)
- · Contamination of the stopping solution

PERFORMANCE CHARACTERISTICS

1. Clinical Performance

A total of 400 specimens from susceptible subjects were tested by a Chinese State Drug Administration (SDA) licensed reference EIA. Comparison for all subjects is showed in the following table:

	RecombiLISA		
Ref. HBc IgM	Positive	Negative	Total
Positive	46	0	46
Negative	0	354	354
Total	46	354	400

Relative Sensitivity: 100%, Relative Specificity: 100%,

Overall Agreement: 100%

WARNING AND PRECAUTIONS

For In Vitro Diagnostic Use

- This package insert must be read completely before performing the test.
 Failure to follow the insert gives inaccurate test results.
- Do not use expired kits.
- 3. Bring all reagents to room temperature (18-28°C) before use.
- Do not use the components of any other type of test kit as a substitute for the components in this kit.
- 5. Do not use serum derived from hemolyzed blood specimens for testing.
- Do not ingest the reagents. Avoid contact with eyes, skin and mouth. Wear protective clothing and disposable gloves while handling the kit reagents and clinical specimens. Wash hands thoroughly after performing the test.
- Do not smoke, drink, or eat in areas where specimens or kit reagents are being handled.
 Users of this test should follow the US CDC Universal Precautions for
- prevention of transmission of HIV, HBV and other blood-borne pathogens.
- Dispose of all specimens and materials used to perform the test as biohazardous waste.
- 10. At the beginning of each incubation and after adding Stop Solution, gently rock the microwells to ensure thorough mixing. Avoid the formation of air bubbles which results in inaccurate absorbance values. Avoid splashing liquid while rocking or shaking the wells.
- Do not allow the microwells to dry between the end of the washing operation and the reagent distribution.
- The enzyme substrate reaction is very sensitive to metal ions. Thus, do not allow any metal elements to come into contact with the conjugate or TMB Substrate.
- The enzyme-substrate is temperature dependent. Ensure that the room temperature for TMB incubation falls between 18-28°C.
- 14. The TMB substrate must be colorless. The appearance of color indicates that the reagent cannot be used and must be replaced. The TMB Substrate must be stored in the dark.
- Use a new dispensing tip for each specimen. Never use the specimen container to distribute conjugate and TMB Substrate.
- 16. The wash procedure is critical. Wells must be aspirated completely before adding the Wash buffer or liquid reagents. Automatic washers must be validated with the test kit prior to use. Insufficient washing will result in poor precision and falsely elevated absorbance values.

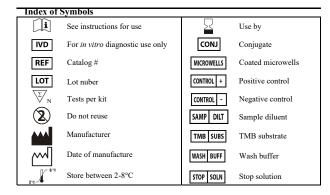
- Microplate reader must be calibrated per manufacturer's instruction to ensure accurate determination of absorbance. Non-calibrated reader often leads to invalid test results.
- Avoid exposure to strong light during color development.

LIMITATION OF THE TEST

- The Assay Procedure and the Assay Result Interpretation must be followed closely when testing the presence of anti-HBc IgM in serum or plasma from individual subjects. Failure to follow the procedure may give inaccurate results.
- The RecombiL/SA HBc IgM ELISA is limited to the qualitative detection of IgM antibody to HBc in human serum or plasma. The intensity of color does not have linear correlation with the antibody titer in the specimen.
- A negative result for an individual subject indicates absence of detectable anti-HBc IgM. However, a negative test result does not preclude the possibility of exposure to or infection with HBV.
- A negative result can occur if the quantity of anti-HBc IgM present in the specimen is below the detection limits of the assay, or the antibodies that are detected are not present during the stage of disease in which a specimen is collected.
- Some specimens containing unusually high titer of heterophile antibodies or rheumatoid factor may affect expected results.
- The results obtained with this test should only be interpreted in conjunction with other diagnostic procedures and clinical findings.

REFERENCES

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