

Pepsinogen I & Pepsinogen II Kit (Microfluidic Fluorescent Immunoassay)

Product name

Pepsinogen I & Pepsinogen II Kit (Microfluidic Fluorescent Immunoassay)

Abbreviated name: LYOFIA PG I & PG II

Ref. No. --- Package size

LMGFPG25C --- 25 Tests, LMGFPG25 --- 25 Tests (N-QC)

Package size

100 Tests, 50 Tests, 25 Tests, 10 Tests, 5 Tests, 100 Tests (N-QC), 50 Tests (N-QC), 25 Tests (N-QC), 10 Tests (N-QC), 5 Tests (N-QC).

Intended use

This device is intended to be used for the in vitro quantitative determination of Pepsinogen I (PG I) and Pepsinogen II (PG II) in human serum or plasma. And it is for professional use only, not for self-testing of untrained individuals, nor for near-patient testing.

Summary

Pepsinogen (PG) is an aspartic protease precursor secreted by the chief cells of the stomach. It is a single-chain polypeptide with a molecular weight of 42kD, which converts into active pepsin in the stomach. PG has seven isoenzymes, which are classified into pepsinogen I (PG I) and pepsinogen II (PG II). PG I is produced by the fundic gland, and PG II is produced by the fundic gland, the cardiac glands, pyloric glands and Brunner's glands. Most of PGs are secreted into the stomach and converts into pepsin to digest food, and about 1% enters the blood circulation. During the atrophy of the fundic gland mucosa, the chief cells of the fundic gland decrease, the secretion of PG I decreases, and the ratio of PG I / PG II decreases. Therefore, the detection of PG I and PG II and the combination of the ratio of the two can screen for atrophic diseases of the fundic gland mucosa. Since atrophic gastritis is related to gastric cancer, so the test can also be used for the auxiliary diagnosis of gastric cancer.

Principle

This product adopts the microfluidic fluorescence immunoluminescence method. The luminescent material relies on the external light source to obtain energy, then it is excited to make luminescence. And the immunological principle used is double antibody sandwich method. In addition, the microstructure in the strip inside the test cassette can make the reaction system to be uniformly mixed inside the test cassette, thereby improving the accuracy and precision of the detection result.

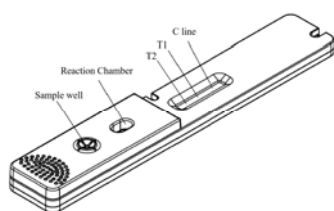


Figure 1: Schematic diagram of the test cassette

As shown in Figure 1, below the sample well is the lyophilized spheres placement tank. The lyophilized spheres are contained in the tank. The main component of the lyophilized sphere is the nanosphere (containing luminescent material) which is coupled with PG I monoclonal antibody I and PG II monoclonal antibody I and DNP-BSA antibody. The main component of T line is PG I monoclonal antibody II and PG II monoclonal antibody II, and the main component of C line is anti-DNP-BSA antibody.

The sample added from the sample well enters the flow microchannel through the microchannel valve and the microchannel mixer valve, so that the lyophilized spheres and the specimen in the lyophilized spheres placement tank are quickly dissolved and mixed evenly. The sample mixture flows along the microfluidic channel to the reaction chamber for reaction. The antigen in the specimen reacts with PG I monoclonal antibody I and PG II monoclonal antibody I to form an

antigen-antibody-nanosphere

complex.

The

antigen-antibody-nanosphere complex will flow forward along the nitrocellulose membrane through the sample pad and can be captured by the PG I monoclonal antibody II and PG II monoclonal antibody II immobilized on the T line of the nitrocellulose membrane to form a double-antibody sandwich complex. In addition, the DNP-BSA antibody in the reaction system can be captured by the anti-DNP-BSA antibody immobilized on the C line. The more antigen in the sample, the more complexes will accumulate on the T line. The intensity of the fluorescent signal reflects the amount of captured antigen.

The fluorescence immunoassay analyzer used with the kit emits emission light, irradiates the T line and the C line, and excites the nanospheres to emit light, and then the specific signal values of the T line and the C line can be obtained.

The content of PG I and PG II in the sample can be determined using the calibration curve served in the Reagent information carrier.

Components and ingredients

No.	Main components and ingredients				
1	Test Cassette	Upper layer microfluidic chip	Fluorescent lyophilized spheres	PG I monoclonal antibody I	
				PG II monoclonal antibody I	
			Blocking lyophilized spheres	DNP-BSA	
		Card shell (containing test strip)	Nitrocellulose membrane	Sample pad	
				PG I monoclonal antibody II	
				PG II monoclonal antibody II	
				anti-DNP-BSA antibody	
			Absorbent paper		
		PVC base plate			
2	Sample Diluent	PBS buffer			
		Proclin300			
3	Reagent information carrier	A calibration curve is stored.			
4	Control	Level 1	PG I antigen and PG II antigen		
		Level 2	PG I antigen and PG II antigen		

Note:

- Sample Diluent (see the packaging label for the quantity);
- The kit whose packaging specifications describes "(N-QC)" do not contain quality control products;
- See the "target value list" for the target value range of the quality controls;
- The components from different lots of kits cannot be interchanged or mixed.

Storage and stability

Store the product at 2~30°C, it has a validity period of 18 months. Once the aluminum foil pouch of the test cassette is opened, the cassette has a validity period of 24 hours. After the control solution is reconstituted, seal and store it at 2~8°C with a validity period of 4 hours. Do not use the test kit beyond the expiration date as indicated on label.

Applicable analyzer

Fluorescence immunoassay analyzer manufactured by Hunan Kangxin Biotechnology Co., Ltd., model LYOFIA-I, LYOFIA8.

Specimen requirements

1. This product is suitable for serum and plasma. Lithium heparin, sodium heparin, EDTA and sodium citrate are the recommended anticoagulants for plasma. The other anticoagulants have not been validated, they may affect the test results.
2. It is recommended that finishing the testing of serum and plasma within 8 hours. If the specimens specified above cannot be used at once, store them at 2~8°C and finish the testing within 7 days, or store the specimens for up to 3 months at -20±5°C.

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3. The samples to be tested should be free of precipitation. If precipitation occurs, centrifugation must be performed first. Do not use heat-inactivated samples.

4. Equilibrate the samples to ambient temperature before measurement. Cryopreserved samples should be completely thawed, rewarmed, and evenly mixed before use. Multiple freeze-thaw cycles should be avoided. Do not use samples with significant hemolysis or blood clots.

Assay procedure

1. Assay preparation

1.1 Please follow this instruction for use and refer to the instruction manual of the fluorescence immunoassay analyzer.

1.2 Turn on the fluorescence immunoassay analyzer, check whether the analyzer can work normally, and prepare other related consumables.

1.3 Equilibrate the aluminum foil pouch to ambient temperature before opening.

1.4 Equilibrate the sample diluent and specimens to ambient temperature.

2. Calibration

Insert the reagent information carrier into the interface for the reagent information carrier on the analyzer LYOFIA-I or LYOFIA8, import the calibration curve stored in the reagent information carrier into the analyzer, and check whether the batch number of the reagent information carrier and the kit are consistent. Refer to the analyzer manual for specific operations.

3. Sample testing

3.1. Take out the test cassette has been equilibrated to ambient temperature and place it horizontally on a flat surface.

3.2. Dilute the sample with the sample diluent at the ratio of 1:3 (recommended procedure: pipette 65μL of sample into a centrifuge tube with 195μL of sample diluent). After mixing, take 65μL of the liquid and quickly add it into the sample well of the upper layer microfluidic chip (the small hole pointed by the arrow on the upper layer microfluidic chip). It is recommended to aspirate and dispense rapidly 3 times in the cassette hole.

3.3. Incubation and testing according to applicable instruments, as follows:

3.3.1. If the measuring instrument is LYOFIA-I, please insert the test card into the incubator immediately after adding the sample and then to let it stand for 10 minutes for reaction. Remove the test cassette after the end of the reaction, and insert it into the right position of the fluorescence immunoassay analyzer LYOFIA-I, click the "Test" for testing, and the analyzer will automatically scan the test cassette.

3.3.2. If the measuring instrument is LYOFIA8, please insert the test card into the test slot immediately after adding the sample, LYOFIA8 will automatically scan the test cassette, time the reaction and automatically detect after the reaction is over.

3.4. The fluorescence immunoassay analyzer automatically detects the results and calculates the content of PG I and PG II in the sample.

3.5. Take out the test cassette used and dispose it as medical waste.

4. Results Analysis

The measured fluorescence signal value can directly read the content of PG I and PG II in the sample from the calibration curve stored in the reagent information carrier of the corresponding batch. The default detection result is in ng/mL.

5. Quality Control

Each laboratory shall establish its own quality control system and rules according to relevant requirements.

To conduct quality control, you must use the quality controls of the same batch of the kit. The quality control product is lyophilized. After returning to ambient temperature, reconstitute it with purified water (show the target list for the water volume required), let it stand for at least 15 minutes, shake it horizontally and mix well, and then test the reconstituted control solution as a sample.

Reference interval

PG I >70 ng/mL; PG I / PG II >3.

It is recommended that each laboratory establish its own reference interval because PG I and PG II level determined is varied

depending upon geographical, individual difference, or testing methods.

Result interpretation

The test results shall be only considered as a clinical reference rather than the unique basis for confirming or excluding a case. For diagnostic purposes, results should always be used in combination with clinical examination, medical history and other results of inspection.

Limitation

1. Possible causes of abnormal test results: Heterophilic antibodies, some non-specific components in blood with similar antigenic determinants can capture fluorescence-labeled antibodies.

2. Bacterial contamination of the sample or repeated freeze-thaw may affect the results.

3. Different brands and materials of blood collection tubes may affect the test results.

Performance characteristics

1. Limit of detection

Items	Limit of detection
PG I	≤2.50 ng/mL
PG II	≤1.40 ng/mL

2. Linearity

Items	Linear interval	Correlation coefficient r
PG I	[3, 200] ng/mL	Not less than 0.9900
PG II	[1.5, 100] ng/mL	Not less than 0.9900

3. Precision

3.1 Repeatability imprecision: The coefficient of variation (CV) is not more than 10%.

3.2 Within-laboratory imprecision: The coefficient of variation (CV) is not more than 10%.

3.3 Inter-lot imprecision: The coefficient of variation (CV) is not more than 15%.

4. Interference: Refer to the method of EP7-A2 "Interference Testing in Clinical Chemistry" to conduct the evaluation. If the relative deviation of the measurand value of the sample spiked with the interfering substance and that of the sample in absence of the interfering substance is not higher than 15%, the substance of no more than the corresponding study concentration may be considered no interference effect. Please see Table 1 for the upper limit of no interference of interfering substances to the assay:

Table 1: List of upper limits of no interference of interfering substances to the assay

Interference substance	Upper limit of no interference to the assay
Triglycerides	10 mg/mL
Bilirubin	0.3 mg/mL
Hemoglobin	6 mg/mL
Rheumatoid factor	50 IU/mL
Heterophilic antibodies	1:10

5. HOOK effect: Please see following table for the concentration that may have HOOK effect.

Table 2: Hook effect

Substance	Hook
PG I	500 ng/mL
PG II	300 ng/mL

Precautions and warnings

1. This product is an *in vitro* diagnostic reagent for single use and must not be reused.

2. The treatment, use, storage of the specimens and kits' each component, and the disposal of solid and liquid wastes generated during the assay process should be handled in accordance with the corresponding measures of local biosafety guidelines or regulations.

3. Strictly follow operation procedure, and the correct result only be obtained under careful operation. Any modification to the operation procedure may affect the accuracy of the test results.

4. This product is sensitive to humidity, do not use if the foil pouch is damaged.

5. Do not insert the test cassette whose surface is wet with other liquids into the analyzer to avoid contamination and damage to the analyzer.

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6. Keep away from vibration and electromagnetic environment when using the test cassette and fluorescence immunoassay analyzer.

7. Please see the outer label of the package of the kit for the production date and expiration date.


⚠ This product contains chemical ingredients. Contacting with skin or mucosa should be avoided. If the product is spilled into eyes, mouth or skin accidentally, rinse with running water and seek for doctor advice if necessary.


⚠ This product contains animal-derived substances. Although it has passed the biosafety test, it does not rule out the risk of other potential infections. Please consider the kit and samples as potential sources of infection, and wear disposable gloves or take other measures to reduce the risk of infection during the detection process.

Symbols for use in the labeling

Symbols	Definition
	KEEP AWAY FROM SUNLPG I & PG IIT
	TEMPERATURE LIMIT
	IN VITRO DIAGNOSTIC MEDICAL DEVICE
	CONSULT INSTRUCTIONS FOR USE
	BATCH CODE
	CATALOG NUMBER
	USE-BY DATE
	DATE OF MANUFACTURE
	MANUFACTURER
	SUFFICIENT FOR TESTS
	DO NOT RE-USE
	CAUTION
	KEEP DRY
	DO NOT USE IF PACKAGE IS DAMAGED
	AUTHORIZED REPRESENTATIVE IN THE EUROPEAN COMMUNITY



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Revision history

Version	Revision date	Change description
V01	2022-05-09	Initial